



## Microspheres: A review on novel approach for enhancement of aqueous solubility of BCS class II drugs

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### Abstract

Oral modified-release multiple-unit dosage forms have always been more effective therapeutic alternative to conventional or immediate release single-unit dosage forms. In terms of the ultimate dosage form, hard gelatin capsules are typically filled with multiparticulates that have been formed into microspheres. Microspheres gained a lot of interest for their ability to target drugs in addition to their sustained release. Multiparticulate drug delivery systems or microspheres, are intended to deliver medications at a particular rate to a specific site. Microspheres are free-flowing powders with particle sizes ranging from 1 to 1000µm that are composed of synthetic polymers or biodegradable proteins. Advantages of using microspheres in industries including bone tissue production, medication delivery, and pollutant absorption and desorption via regeneration. The study demonstrates the microsphere parameter design and measurement process. There are several different types of microspheres, including bioadhesive, polymeric, magnetic, floating, and radioactive microspheres. Microspheres have potential applications in multiple domains, including gene delivery, ocular drug delivery, target drug delivery, oral drug administration, and cosmetics, among others mentioned in the paper. Achieving the best therapeutic efficacy requires delivering the drug to the target tissue at the proper concentration in a specific duration of time, with the least amount of toxicity and side effects possible. There are various methods for continuously managing the delivery of the drug to the intended site. Using microspheres as drug carriers is one such approach. The current aim of this review is to investigate the formulation, assessment, and characterization processes as well as other aspects of the microparticulate drug delivery system in a constant, controlled manner.

**Keywords:** Microspheres, controlled release, novel drug delivery, method of preparations

### Introduction

The majority of the time, it is observed that the *in vitro* data do not correlate with the *in vivo* data. This is primarily due to factors such as inadequate or poor absorption, fast metabolism and elimination (such as peptide drugs), drug distribution to related tissues (such as cancer drugs), low aqueous solubility of drugs, high fluctuations in drug plasma levels caused by unpredictable bioavailability following oral administration, and the impact of food on plasma levels. A promising approach to overcome the previously described challenges is the development of appropriate drug delivery systems that have an ability to release the active ingredient in accordance with the particular needs of the therapy being administered. In addition to combining the benefits of colloidal drug carrier systems including liposomes, polymeric nanoparticles, emulsions, and solid lipid nanoparticles (SLN) also overcomes the disadvantages of these systems [1].

Microcapsules refer to those in which the entrapped material is clearly surrounded by the characteristic capsule wall, while in micromatrices the entrapped material is distributed throughout the matrix of the microsphere. The drug has been incorporated into the solid biodegradable microspheres and disseminated or dissolved by the particle-matrix. These microspheres have the potential to provide controlled drug release. Solid spherical particles with dimensions ranging from 1 to 1000µm are called microspheres. These are spherical, freely-flowing, biodegradable particles composed of synthetic polymer or proteins [2]. Microspheres are divided into two types:

- Microcapsules
- Microstructures

Having solid biodegradable microspheres containing a medicine dissolved or dispersed via a particle matrix enables the controlled release of a medication. They are composed of polymeric, waxy, or other protective components, as well as biodegradable synthetic polymers and modified natural goods. Polymers and waxes composed of both natural and synthetic components are used in their production. The solubility, drug release and stability of microspheres are influenced by the type of polymer that is utilised to produce them. Expandable, polystyrene, and polyethylene microspheres are the most widely used types of polymeric microspheres. Microspheres can be solid or hollow in nature. They are used to lower the density of a material. Topical formulations based on microspheres have become more popular due to their long-lasting therapeutic effects. In recent years, the usage of microparticles as a carrier for drug delivery has been increased [3, 4, 5, 6, 7].

A wide range of fundamental components have been encapsulated, such as adhesives, agrochemicals, living cells, active enzymes, flavours, scents, vitamins, water, and medications. This review provides brief description on the benefits, history, evaluation techniques, preparation strategies, and the most recent advancements and applications [8].

### Advantages of microparticles [9]

Because of their distinct characteristics, such as size and shape, MDDS has numerous advantages, among which are explained in brief as follows:

- They provide the drug with protection from the gastrointestinal tract and other external surroundings

while also assisting in the controlled and prolonged release of the enclosed drug.

- MDDS contributes to the improvement of therapeutic benefit and reduction of adverse effects by enhancing the solubility characteristics of poorly soluble drugs.
- MDDS reduces toxicity concerns and enhances patient compliance by reducing the need for repeated drug administration and by disguising the taste and odour of medications (for example solfa medicines and fish oils).
- MDDS additionally facilitates the delivery of incompatible agents by encapsulating them within a single shell.
- The encapsulation of living cells (sealed erythrocytes).
- Converting a liquid into a solid that flows freely.
- Careful handling of hazardous substances
- Makes it easier for substances that are insoluble in water to dissolve in aqueous media.
- Immunisation transition and gene therapy employ pH-triggered microparticles.
- One advantage of parenteral microparticles is that they can deliver high doses of water-soluble medications without causing severe osmotic responses at the administration site.
- Enhanced solubility, dispersibility, and flow ability.

#### Disadvantages of microparticles <sup>[9]</sup>

- It is regulated by several factors, including the type of polymer, residency duration, and the presence of food.
- The active ingredient's release kinetics vary depending on the dose at which it is developed.
- Drug delivery may occasionally occur at a location different than the intended site due to polymer breakdown.
- The development of MDDS is also significantly more expensive than developing comparable conventional preparations.
- High standardisation of the process is required to achieve reproducibility of the formulations.
- Process variables that influence the stability of the core particles to be encapsulated include temperature changes, pH changes, solvent additions, evaporation, and agitation.
- The structure of the microparticle manufacturing process can be altered, and different techniques including targeted delivery, intracranial local delivery, for neurological diseases, and others can be employed to get around these limitations.

#### Types of Microspheres

##### Bioadhesive microspheres

The technique of adhering drug to the membrane by means of water-soluble polymers with adhesive properties is known as adhesion. The attachment of a system that delivers drugs to a mucosal membrane, such as the nasal, ocular, buccal, or rectal membranes, is referred to as bio adhesion. Because these microspheres stay at the application site for a longer duration of time, and interact with the site of absorption and have better therapeutic effects <sup>[10]</sup>.

##### Magnetic microsphere

The ability to deliver the medication precisely to the required site makes this kind of delivery method crucial. In this condition, a higher amount of freely circulating drug will be replaced by a smaller amount of magnetically

focused drug. Magnetic microspheres are comprised of dextran, chitosan, and other incorporated materials respond magnetically to a magnetic field. The various types consist of:

##### a. Therapeutic Magnetic microspheres

These are used to treat liver tumours using a chemotherapeutic drug. This technique can also target drugs such as proteins and peptides <sup>[10]</sup>.

##### b. Diagnostic microspheres

By producing nano-size particles known as paramagnetic iron oxides, they can be used to distinguish bowel loops from other abdominal structures and to image liver metastases <sup>[12]</sup>.

##### Floating microspheres

Floating forms do not affect the rate of gastric emptying because their bulk density is lower than that of gastric fluid; they float in the stomach. The drug is released gradually and at the appropriate rate when the system is floating on gastric content, which enhances the variability of plasma concentration and stomach residency. There is also reduced chance of strikes and dose dumping. Additionally, its therapeutic effect lasts longer, resulting in fewer dose intervals <sup>[13]</sup>.

##### Radioactive microspheres

Radioembolization therapy with 10–30 nm microspheres size greater than capillary microspheres, are inserted into the arteries that produce an interest tumour by tapping into the first capillary bed as they pass through. Therefore, in each of these scenarios, radioactive microspheres deliver a substantial radiation dose to the intended regions while sparing the surrounding normal tissues. Unlike drug delivery systems, it operates from a radioisotope-typical distance rather than releasing radioactivity from the microspheres. The three types of radioactive microspheres are  $\alpha$ ,  $\beta$ , and  $\gamma$  emitters <sup>[14]</sup>.

##### Polymeric microspheres

The different types of polymeric microspheres can be classified as:

##### Biodegradable polymeric microspheres

Because natural polymers like starch are biodegradable, biocompatible, and bioadhesive, they are utilised. Biodegradable polymers have a high degree of swelling in aqueous media, which allows them to stay in touch with mucous membranes for an extended period of time and form gels. The rate and extent of medication release are controlled by the polymer concentration and the release pattern throughout time. The primary drawback is the complex drug loading that occurs with biodegradable microspheres in clinical applications, which makes controlling of drug release challenging. However, they have a wide range of uses in microsphere-based therapy <sup>[15]</sup>.

##### Synthetic polymeric microspheres

It has been demonstrated that synthetic polymeric microspheres are safe and biocompatible, and they are frequently utilised in clinical applications as bulking agents, embolic particles, drug delivery vehicles, fillers, and other applications <sup>[16]</sup>. The primary drawback of these

microspheres is their propensity to disperse from the injection site, which increases the possibility of embolism and additional organ injury.

### Mechanism of Microspheres

Most drug delivery systems using microparticles inhibit the development of an internal solid dispersion morphological structure resembling a matrix. The drug is released through erosion and might not dissolve in the polymeric matrix. Water first permeates the matrix and dissolves the resultant material close to the device's surface. By creating a passageway to the surface and delivering an appropriate amount of drug in the initial drug burst, the subsequent osmotic pressure is reduced [17].

#### a. Degradation-controlled Monolithic System

The drug dissolves in the matrix and is distributed uniformly throughout the degradation-controlled monolithic system. It fuses to the matrix and is only released when the matrix breaks down. The medication diffuses more gradually than the matrix breaks down.

#### b. Diffusion-Controlled Monolithic System

In this system, the degradation of the polymer matrix occurs concurrently or prior to the release of the active ingredient through diffusion. The frequency of release is also influenced by the type of degradation process used by the polymer.

#### c. Diffusion-Controlled Reservoir System

In the diffusion-controlled reservoir system, the active substance is enclosed by a rate-controlling membrane that allows it to diffuse. The membrane erodes until delivery is accomplished. During this process, matrix degradation has a minor impact on drug release [18].

#### d. Osmosis

The microcapsule's polymer coating acts as a semipermeable barrier during osmosis, causing an osmotic pressure differential between the inside and outside of the microcapsule. The medication solution can therefore be pulled out of the microcapsule through microscopic pores in the covering [19].

#### e. Erosion

Drug release happens when certain coating materials, such as glyceryl mono stearate, beeswax, and steryl alcohol, undergoes coat erosion as a result of pH and enzymatic hydrolysis.

### Materials and Methods

#### Materials [20]

Materials such as polymers are typically used to produce microspheres. They are classified into two types.

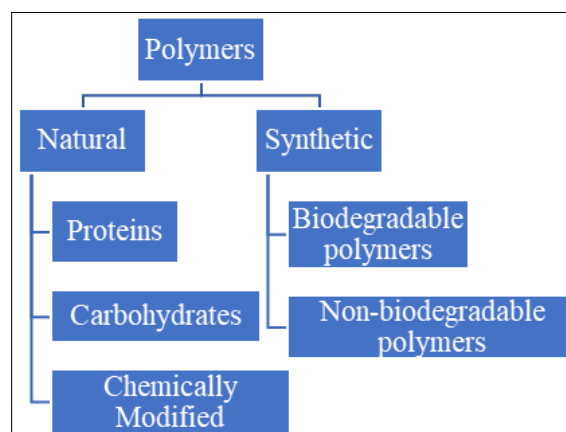


Fig 1: Polymers used in Microspheres Development

Synthetic polymers are divided into two types.

#### 1. Non-biodegradable polymers

- Acrolein
- Poly methyl methacrylate (PMMA)
- Epoxy polymers
- Glycidyl methacrylate

#### 2. Biodegradable polymers

- Poly anhydrides
- Lactides, Glycolides & their co polymers
- Poly alkyl cyano Acrylates

Natural polymers obtained from numerous sources such proteins, carbohydrates and chemically modified carbohydrates.

#### 1. Proteins

- Collagen
- Albumin
- Gelatin

#### 2. Carbohydrates

- Carrageenan
- Chitosan
- Agarose
- Starch

#### 3. Chemically modified carbohydrates

- Poly starch
- Poly dextran

**Solvents:** Most common solvents employed as oil phase for the preparation of MPs are Dichloromethane, Chloroform, Tetrahydrofuran, Ethanol, Isosorbide dimethyl ether, N-Methylpyrrolidone, Ethyl acetate, DMSO, Acetone, Methyl ethyl ketone, Propylene carbonate, Ethyl propionate, Acetonitrile etc.

#### Criteria For Microsphere Preparation

The micro encapsulation technique can be employed to incorporate solid, liquid, or gaseous materials into one or

more polymeric coatings [21]. The development of distinct microspheres using different methods is dependent on factors such as particle size, administration route, drug release duration, and these characters connected to rpm, method of cross-linking, evaporation time, coprecipitation, etc [22]. The following requirements should be accomplished for preparation of microspheres [17]

1. The capacity to load drug in reasonably high doses.
2. The preparation's stability following synthesis and its shelf life, which is therapeutically acceptable.
3. Regulated particle size and dispersibility in injectable aqueous media.
4. Well-controlled, broad-spectrum release of the active reagent.
5. Biodegradability under control biocompatibility.
6. Openness to alterations in composition.

### Methods of Preperation

Methods used for the preparation of microspheres are

- Single emulsion techniques
- Double emulsion techniques
- Polymerization
- Normal polymerization
- Interfacial polymerization
- Phase separation coacervation technique
- Spray drying
- Emulsion-Solvent diffusion technique
- Solvent evaporation
- Ionic gelation method

### Single emulsion technique [2]

This procedure is used for the preparation of various proteins and carbohydrates. The natural polymers first undergo dissolution in an aqueous phase and then distributed in a non-aqueous phase known as the oil phase. That is the first step in the procedure. The next step is cross-link which is done by using two methods:

#### Cross-linking by heat

In this method dispersion is added into heated oil, yet this method is inappropriate for drugs that are thermolabile.

#### Chemical cross-linking agents

by employing of chemicals such as glutaraldehyde, formaldehyde, and diacid chloride etc. However, when applied during preparation and then centrifuged, cleaned, and separated, it is harmful to the unwanted exposure of active components to contaminants. Apply w/o emulsion to the liquid paraffin with a surfactant to dissolve the chitosan solution (in acetic acid). Glutaraldehyde in a 25 percent solution is used as a cross-linking agent to create microspheres.

### The double emulsion technique [23]

This process is most appropriate for water-soluble drugs, proteins, vaccines, and peptides. It produces multiple emulsions or double emulsions of the type w/o/w. This method works with both natural and synthetic polymers. The aqueous protein solution is distributed in the lipophilic organic continuous phase. The active ingredients might be present in this protein solution.

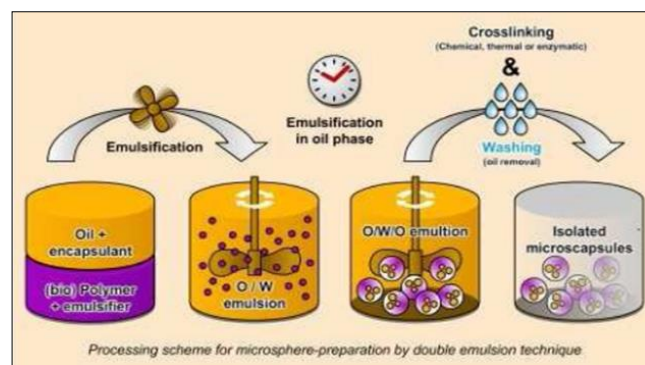


Fig 2: Microspheres by Double Emulsion Technique [20]

### Polymerization techniques

Following two techniques are mainly used for the formulation of microspheres

#### Normal polymerization

In bulk polymerization, the polymerization process is typically started by heating a monomer, or a mixture of monomers, the catalyst, and the initiator. The resulting polymer can be formed as microspheres. Addition of the drug during the polymerization process is one way to accomplish this. Although it is a pure polymer production method but the heat generated by the reaction is particularly difficult to dissipate, which damages the thermolabile active components. In suspension polymerization, also known as pearl polymerization, the active drug is dissolved in droplets in a continuous aqueous phase through heating of the monomer combination. This process is done at a lower temperature.

#### Interfacial polymerization

The dispersed phase is effectively enveloped by a polymer film that is created by the interaction of different monomers at the interface between the two immiscible liquid phases. Two reactive monomers are used in this approach; first one is by dissolved in the continuous phase, and the other is distributed in an aqueous continuous phase, which is where the second monomer is emulsified [2].

#### Phase separation coacervation technique

The concept behind this technique is to influence the production of a polymer-rich phase known as coacervates by reducing the polymer's solubility in the organic phase. This procedure involves dispersing the drug particles into a solution of polymers and then adding an incompatible polymer to the device, which causes the initial polymer phase to separate and swallow the drug particles. The polymer solidifies as a result of adding the non-solvent. Butadiene has been utilised as an incompatible monomer in this technique to create microspheres of polylactic acid (PLA). Process factors are important because the dispersion of the polymeric film is determined by the coacervate accomplishment rate, the particle size and how they aggregated to produce larger particles. Agglomeration must be prevented by employing a speed-appropriate stirrer to mix the suspension, since the produced polymerized globules will start to attach and form agglomerates as soon as the microsphere-forming process begins. Consequently, since there is no specific state of equilibrium attainment, process factors are crucial because they control the kinetics of the produced particles [3].

### Spray drying and spray congealing method

The spray drying and spray congealing methods differ based on whether the solvent is eliminated or the solution is cooled. Both the drug and the polymer are homogenised in a high-speed homogenizer after being dispersed in an organic volatile solvent. The resultant dispersion is then sprayed via a hot air stream, which causes the solvent to rapidly evaporate and forms the microparticle. The microparticles are separated from the heated air using a cyclone separator, and any leftover solvent is then removed by vacuum drying. Important advantages of this method include high encapsulation efficiencies and the lack of surfactant residue upon the surface of the microparticles. Microparticle size and shape are influenced by a number of variables, including temperature, pressure, nozzle diameter, mixture volume, polymer and drug concentration, feed flow rate, and inlet temperature. In general, flavours, oils, and perfumes are encapsulated via spray drying, an inexpensive industrial technique. This method's main objectives are to safeguard thermolabile active components and stop delicate medications from oxidising. By using several drying processes, it enhances product quality and guards against environmental damage to the product. The dispersion of the active ingredient in heated coating material without the use of a solvent is called spray congealing, or spray chilling. After that, the mixture is sprayed onto a stream of cold air, where it cools to a temperature below the coating material's melting point, forming solid droplets.

### Emulsion-Solvent Diffusion Technique

Using the emulsion solvent diffusion approach, floating microparticles were produced in order to increase the residence period in the colon. After dissolving the drug polymer mixture in a 1:1 ratio of ethanol and dichloromethane, the mixture was gradually added to a solution of sodium lauryl sulphate (SLS). For one hour, the solution was agitated at room temperature at 150 rpm using a propeller-style agitator. As a result, the generated floating microspheres were cleaned and allowed to dry at room temperature in a desiccator. The subsequent microparticles were collected and sieved [20].

### Solvent evaporation/ extraction method

Oil-in-water (O/W) emulsification is the primary method of encapsulating hydrophobic drugs through solvent extraction. The quick hardening period and the active ingredient's immediate integration into the microparticles are this method's primary benefits. The solvent extraction or evaporation process is completed by dissolving the drug component and polymer in a suitable organic solvent. Stirring the solution of Surfactant produces an emulsion. The MPs are eventually gathered when the solvent has evaporated. It is possible to produce the hollow interior core of the floating microparticle dosage form by employing evaporation and solvent diffusion methods for the development of such systems, polymers such as cellulose acetate, chitosan, Eudragit, Acrycoat, Methosyl, polyacrylate, polyvinyl acetate, Carbopol, agar, polyethylene oxide, and polycarbonate have all been studied. Compact microparticles arise from high-temperature solvent evaporation from the emulsion or solvent extraction with a large amount of water. The two most popular organic solvents utilised in emulsion solvent evaporation procedures are dichloromethane and chloroform [9].

### Ionic gelation method

The drug is introduced to an aqueous sodium alginate solution at this stage of the procedure and stirring continuously. The resulting microspheres were allowed to internalise for a full day in the original solution before being filtered to facilitate separation. The drug will not release at an acidic pH; complete release occurs at pH 6.4–7.2 [17].

### Evaluation parameters of Microspheres

#### Characterization

The identification of the microparticulate carrier is an important phenomenon that helps in the development of an appropriate carrier for the delivery of antigens, drugs, or proteins. The microspheres exhibit a range of microstructures. These microstructures control the carrier's stability and release [20].

#### Particle size determination [9]

The MP's particle size and particle size distribution were ascertained through the application of optical microscopic technique. The stage micrometre was used to calibrate the eyepiece micrometre. A small amount of MPs were dispersed over a clean glass slide. To determine the particle size, about 100 particles from each batch were assessed. Differential scanning calorimetry analysis can be used to determine crystallinity.

The two most popular methods for visualising microparticles are scanning electron microscopy (SEM) and conventional light microscopy (LM). Both are useful for examining the external structure and form of microparticles. For double-walled microspheres, LM offers control over the coating parameters. Before and after coating, the microspheres structures may be observed, and the change can be quantified under a microscope. Comparing SEM to LM, a better resolution is offered. SEM makes it possible to examine the microspheres' surfaces and, in the event that the particles are cross-sectioned, to examine structures with two walls.

#### Flow properties [2]

##### Density:

##### Bulk density

It is calculated by measuring the length of a measuring cylinder after adding a sample of known-weight microspheres without tapping, and then dividing the weight by the volume.

$$\text{Bulk density} = \frac{\text{wt. of microspheres}}{\text{bulk volume}}$$

##### Tapped density

It is determined by adding a known-weight sample of microspheres to a measuring cylinder, and tapping it and then measuring its volume, and then density is calculated by using formula;

$$\text{Tapped density} = \frac{\text{wt. of the microspheres}}{\text{volume after tapping}}$$

##### Hausner's ratio

Microparticle flow can be predicted using Hausner's ratio. A free-flowing microparticle can be identified by a Low Hausner's ratio of less than 1.2.

Hausner's ratio = bulk density – tapped density

**Angle of repose**

It is described as the greatest angle that a pile of microspheres can make with respect to the horizontal. The methods for figuring out the angle of repose are the fixed height cone and the fixed base cone.

$$\text{Angle of Repose } \theta = \tan^{-1} h/r$$

r = the radius of the base of the heap of microsphere

h = height of the heap of microsphere

**Zeta potential (ZP) measurement**

Using the Malvern Zetasizer Nano ZS 90 (Malvern Instruments, UK), the electrophoretic mobility was determined in order to measure the zeta potential. A field strength of 20 V cm<sup>-1</sup> was applied. All samples were diluted in distilled water before testing.

**Percentage yield**

The percentage yield was determined by using the formula given as:

$$\text{Percentage yield} = \frac{\text{Practical yield of drug (gm)} \times 100}{\text{Wt. of drug taken (gm)}}$$

**Drug entrapment efficiency**

Drug-containing microspheres (5 mg) are crushed and dissolved in distilled water for three hours with the aid of an ultrasonic stirrer, filtered, and then subjected to UV-visible spectroscopic analysis.

$$\text{Entrapment efficiency} = \frac{\text{actual drug content}}{\text{theoretical drug content}}$$

**Swelling index**

The swelling index of the microsphere was determined by using the formula,

Swelling index = mass of swollen microspheres - mass of dry microspheres/mass of dried microspheres

**In vitro drug release study**

900 ml of phosphate buffer of pH 7.4 used as the dissolving medium for eight hours during the drug release study, which is conducted using a USP dissolution test apparatus paddle type at 37 ± 0.5°C and 100 rpm. For the test, microspheres containing 10 mg of drug are utilised. At specified intervals, 5 ml of the sample solution was removed, filtered, appropriately diluted, and subjected to spectrophotometric analysis at an appropriate wavelength. As soon as the test sample was removed, the same volume of new dissolving medium was added.

**Stability studies**

Can be done by placing the microspheres in screw capped glass container and stored them at following conditions:

1. Ambient humid condition
2. Room temperature (27±/-2 0C)
3. Oven temperature (40±/-2 0C)
4. Refrigerator (5 0C -80C)

The study was carried for 60 days, and the drug content of the microsphere was examined.

**Fourier Transform-Infrared Spectroscopy**

Fourier Transform Infrared Spectroscopy can be used to assess the carrier system's polymeric matrix deterioration. ATR, or altered total reflectance, is used to measure the microspheres' surface. The infrared (IR) spectra of the surface material were mostly obtained from many reflections of the IR beam that passed through the alternating total reflectance cell within the sample. The alternating overall reflectance-Fourier Transform-Infrared Spectroscopy gives insight into the surface composition of the microspheres based on the manufacturing processes & conditions.

**Applications of Microspheres****Microspheres in vaccine delivery**

The following requirements must be satisfied by the perfect vaccine: affordability, ease of use, safety, and effectiveness. Conventional vaccinations have limitations that may be addressed by biodegradable vaccine delivery systems for parenterally delivered vaccines [2].



**Fig 3:** Therapeutic effects and advantages of MDDS in vaccine delivery<sup>9</sup>

**Monoclonal antibodies mediated microspheres targeting [2]**

Monoclonal antibodies targeted at microspheres are known as immune microspheres. This is a method for reaching certain sites with targeted targeting. High specificity molecules are called monoclonal antibodies. Direct attachment of Mabs to the microspheres is made possible by covalent binding. Connecting Mabs to microspheres can be done using any of the techniques listed below.

- Adsorption that is both selective and non-specific;
- direct coupling;
- and coupling through reagents

**The microspheres in chemotherapy**

Utilising microspheres as carriers for anti-tumour drugs is one of their most exciting applications. The administered microspheres caused leaky vasculature and increased endocytic activity. The soluble polyoxy ethylene is coated to prepare the stealth microspheres. The build-up of non-stealth microspheres in the RES [Reticulo Endothelial System] can also be used for cancer treatment [23].

**Biomedical Therapeutic [9]**

A wide range of medicinal and biological research applications utilises microparticles

- Porous silicon microparticles used to deliver therapeutic siRNA
- Using microcapsules or nanocapsules based on chitosan to transport drugs, biologicals, and vaccines.
- Using intracellular agents-loaded microparticles for cell engineering, cellular phenotyping can be controlled.
- Alveolar macrophages are useful for solid lipid microparticle inhalation in tuberculosis.
- Microparticles of gentamicin and dextran to treat bacterial wound infections.
- Thiolated polydimethyl aminoethyl methacrylate (PDCy) synthesis and *in vitro* utilisation of PDCy submicroparticles as oral insulin carriers.

**Biosensors**

Numerous recent research has examined the application of core-shell particles as biosensors in biomedical fields such as medical diagnostics. Implantable biosensors are in high demand because they allow for real-time monitoring and early diagnosis of a variety of disorders through continuous monitoring and analyte detection *in vivo*. Biomarkers including glucose, enzymes, ions, DNA, and antibodies can be found using biosensors that contain bio sensing agents and signalling factors. The development of intelligent microparticles with intricate architectures has made it possible to construct biosensors with a wider range of applications thanks to microfluidics. A hollow polyethylene glycol microcapsule was developed by Xie *et al.* To contain nanosensors in a liquid core. Implantable biosensors are used *in vivo* to identify biomolecules that are confined by movement or released from their original place. A potential way of dealing with this problem is to embed the nanosensors within a hydrogel scaffold. But the operation of the sensor can be interfered with if nanosensors come into touch with the hydrogel wall. Thus, in order to keep the

nanosensors from coming into touch with the hydrogel, the scientists created a microcapsule with a microfluidic liquid core. Heparin-sensitive gold nanorods, core-encapsulated gold nanorods, and glucose-sensitive quantum dots are examples of nanosensors. Biomolecules that travel through the envelope, such heparin and glucose, interact with nanosensors to produce optical signals that can be detected [9].

**Molecular Imaging**

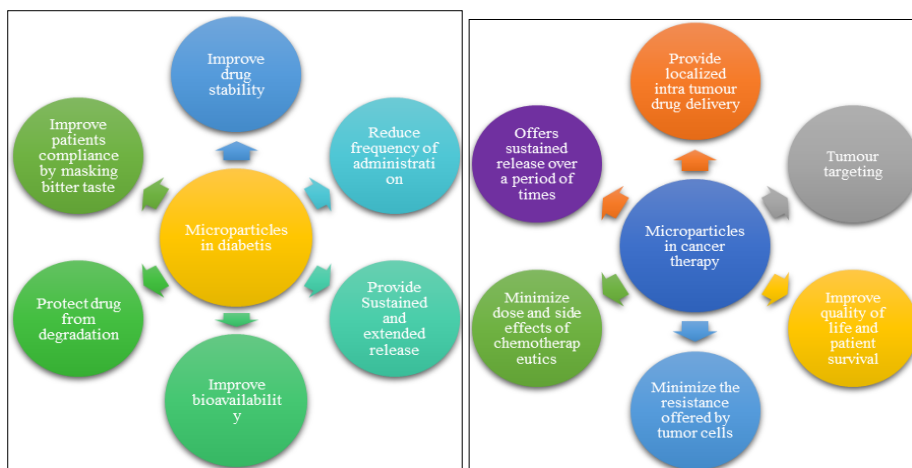
The microspheres have been utilised for targeting and have undergone substantial research. Radiolabelled microspheres can be used for imaging a variety of cells, cell lines, tissues, and organs. When imaging specific areas, the microspheres' spectrum of particle sizes plays a crucial role. The intravenous particles will become caught in the lung's capillary bed if they are injected somewhere other than the portal vein. This phenomenon is used with tagged human serum albumin microspheres for scintigraphic imaging of lung tumour masses [20].

**Targeting using Microparticulate Carriers**

Targeting, or site-specific medication delivery, is a well-established idea that is receiving a lot of attention. The drug's ability to specifically engage and gain access to its target receptors determines how effective it is as a treatment. The main mechanism of pharmacological action, which is mediated by the employment of a carrier system, is the capacity to exit the pool in a repeatable, effective, and targeted manner. The physical characteristics of the surroundings or the particles' biophysical interaction with the target tissue's cells cause the particles to be retained when they are placed in discrete anatomical compartments.

**In Oral drug delivery**

Drugs intended for oral use typically rely on how well they dissolve and absorb. Due to their tiny size, these medications have a higher oral absorption and bioavailability while having a poor water solubility and low bioavailability. Completely absorbed but slowly acting medications have a quick acting form because of microparticles. The microparticles were made with Eudragit RS 100 and cellulose acetate. After 12 hours, the drug release from the microscopic particles was measured microparticle floating oral medication delivery system may work well for cardiac delivery in place of traditional oral tablets [9].



### Recent Advancement in Microsphere<sup>[20]</sup>

#### ▪ Important utilizations of chitosan polymer Cholesterol-lowering effects

Examples of fibres having high, moderate, and low bile acid-binding capabilities were chitosan and cellulose, respectively. When either of these fibres were included in the diet at a rate of 7.5%, the blood cholesterol levels in the control group of mice given an excessive fat/high cholesterol diet for three weeks increased by about two times to 4.3mM. Furthermore, the administration of these fibres decreased the quantity of cholesterol that the HFHC diet had caused to build up in the liver's reserves. The hypocholesterolaemic action of the three types of fibres was comparable, however cholestyramine caused the highest reduction in liver tissue cholesterol levels. The three main processes that underlie cholestyramine's ability to lower cholesterol are:

1. reduced dietary intake of cholesterol;
2. decreased absorption of cholesterol; and
3. increased excretion of cholesterol and bile acid from the faeces. The latter effects are explained by cholestyramine's strong bile acid-binding ability. Conversely, adding chitosan or cellulose to the meal decreased the amount of food that contained cholesterol, but it had no effect on the absorption of cholesterol in the intestinal tract or the production of faecal sterol. The current study offers compelling evidence that the satiety and satiation effects are the primary cause of the LDL lowering.

#### ▪ Increase Stability of Drug

In order to generate a slurry and knead the drug for 45 minutes till dough mass, chitosan polymer is utilised to boost the stability of the drug. The resulting dough mass is sent through sieve number 16, which creates granules that are entirely stable under various circumstances.

#### ▪ Orthopaedic Patients

Because of its osteoconductive, wound-healing-enhancing, and antibacterial qualities, chitosan is a biopolymer that is appealing for use as a bioactive covering to increase the osteointegration of orthopaedic and craniofacial implant devices. It has been demonstrated to be helpful in expediting bone regeneration and wound healing as well as encouraging tissue growth in tissue repair.

#### ▪ Cosmetics industry

Cosmetic compositions with the presence of novel quaternary derivatives of chitosan of the formula are made available for the purpose of treating of hair or skin. The chitosan derivatives exhibit good significant properties, especially with regard to hair keratin, and demonstrate conditioning and strengthening properties for hair. For example, oxidation and hair setting lotion Compositions of Skin Cream, Hair Treatment, Hair Colouring, Hair Toning, and Gel Form.

#### Future Challenges

Because of its wide range of applications in molecular biology, including the detection of six single nucleotide polymorphisms using a genotyping platform, the prevention of tumours following liver transplantation using yttrium-90 microspheres, and the highly advanced method of delivering vaccines and proteins, the future of microspheres appears bright.

### Conclusion

Because microspheres have better patient compliance and targeting accuracy than other medication delivery technologies, they are safer to use. Due to its advantages in terms of bioavailability, lower dose frequency, improved stability, dissolution rate, and continuous and controlled-release action, the microsphere drug delivery system is the most widely used drug delivery method. An effective and safe drug delivery method that may be applied to many different tasks, including precision medication targeting, floating and vaccination delivery, is the microsphere drug delivery system. There are numerous, efficient methods for preparing and assessing microspheres that have been developed. In addition to being used for delivering drugs, microspheres are also employed in cancer treatment, tumour imaging, and biomolecular interaction detection. Microspheres will therefore become more significant in the field of medicine in the future.

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### References

1. Chalikwar SS, Belgamwar VS, Talele VR *et al.* Formulation and evaluation of nimodipine loaded solid lipid nanoparticle delivered via lymphatic transport system. *Colloid Surf B*,2012;97:109–116.
2. Raj H, S Sharma, A Sharma, K Verma, A Chaudhary. "A Novel Drug Delivery System: Review on Microspheres". *Journal of Drug Delivery and Therapeutics*,2021;11(2-S):156-61. doi:10.22270/jddt.v11i2-S.4792.
3. Jain NK. Controlled and Novel drug delivery; CBS Publishers New Delhi, India; 04 Edition,2004:21:236-237.
4. Venkatesan P, Manavalan R, Valliappan K. Microencapsulation: a vital technique in novel drug delivery system. *Journal of Pharmaceutical Sciences and Research*,2009;1(4):26-35.
5. Ramteke KH, Jadhav VB, Dhole SN. Microspheres: as carriers used for novel drug delivery system. *IOSR Journal of Pharmacy*,2012;2(4):44-8.
6. Chein YM. Novel Drug Delivery System, second edition, revised & expanded, Marcel Dekker, Inc, New York, 1992.
7. Sachan AK, Gupta A, Arora M, Formulation & characterization of nanostructured lipid carrier (NLC) based gel for topical delivery of etoricoxib, *Journal of drug delivery and therapeutics*,2016;6(2):4-13.
8. M Suresh Babu, Arkaan Qamar Abbas. Microencapsulation: A Review, *European Journal of Biomedical and Pharmaceutical Sciences*,2019;6(6):224- 237.
9. A review Microparticulate drug delivery system. Sandhya Gujare, Pallavi Jadhav, A. H. Hosmani,2023;8(2):1287-1303.
10. Liu G, Yang H, Zhou J, Preparation of magnetic microsphere from water-in-oil emulsion stabilized by block copolymer dispersant. *Biomacromolecules*,2005;6:1280-1288.

11. Shanthi NC, Gupta R, Mahato KA. Traditional and Emerging Applications of Microspheres: A Review, International Journal of Pharm Tech Research,2010;2(1):675-681.
12. Najmuddin M, Ahmed A, Shelar S, Patel V, Khan T. Floating Microspheres of Ketoprofen: Formulation and Evaluation, International Journal of Pharmacy and Pharmaceutical sciences,2010;2(2):83-87.
13. Hafeli U. Physics and Chemistry Basic of Biotechnology. Focus on biotechnology. Review. Radioactive Microspheres for Medical Application,2002;7:213-48.
14. Saralidze K, Leo H, Koole, Menno L, Knetsch W. Polymeric Microspheres for Medical Applications, Materials,2010;3:3357-3564.
15. Trivedi P, Vermal, Garud N. Preparation and Characterization of Aceclofenac Microspheres, Asian Journal of pharmaceutics,2008;2(2):110-115.
16. Nikam VK, Gudsoorkar VR, Hiremath SN, Dolas RT, Kashid VA. Microspheres-A Novel drug delivery system: An overview; International Journal of Pharmaceutical and chemical sciences,2012;1:113-128.
17. Alagusundaram M, Madhu C, Umashankari K, Attuluri B, Lavanya C, Ramakant S. Microspheres: As novel drug delivery system. International Journal of Chem Tech Research,2009;1(3):526-534.
18. Nitika A, Ravinesh M, Chirag G, Manu A. Microencapsulation, A novel Approach in Drug Delivery. A review, Indo Glob. J. Pharm. Sci,2012;2(1):1-20.
19. Brazel CS, Peppas NA. Modeling of drug release from swellable polymers. European journal of pharmaceutics and biopharmaceutics,2000;49(1):47-58.
20. Microspheres as drug delivery system-a review BS Prasad, VR Gupta, N Devanna, K Jayasurya J Glob Trends Pharm Sci,2014;5(3):1961-72.
21. Ghulam M, Mahmood A, Naveed A, Fatima RA. Comparative study of various microencapsulation techniques. Effect of polymer viscosity on microcapsule characteristics, Pak.J.Sci,2009;22(3):291-300.
22. Li SP, Kowalski CR, Feld KM, Grim WM. Recent Advances in Microencapsulation Technology and Equipment, Drug Dev Ind Pharm,1988;14:353-376.
23. Patil NV, Wadd NV, Thorat SS, Sudarshan US. Microspheres: A novel drug delivery system. Am. J. PharmTech Res,2020;10(02):286-301.
24. Bhalekar M, Upadhaya P, Madgulkar A. Formulation and characterization of solid lipid nanoparticles for an anti-retroviral drug darunavir. *Appl Nanosci*,2017;7:47-57.
25. Yadav AV, Mote HH. Development of Biodegradable Starch Microspheres for Intranasal Delivery, Indian Journal of pharmaceutical Sciences,2008;70(2):170-174.
26. Gupta AK, Dey BK. Microencapsulation for controlled drug delivery: a comprehensive review. Sunsari technical college journal,2012;1(1):48-54.
27. Sachan NK, Singh B, Rao KR. Controlled drug delivery through microencapsulation. Malaysian J Pharm Sci,2006;4(1):65-81.
28. Desai Ujwala, Bhavsar Dhaval. Formulation and Evaluation of Microparticles Formed by Amphiphilic Crystallization Technique: Optimization of Process Parameter. World Journal of Pharmaceutical Research,2014;3:1025-1054.
29. Priyadarshini Soni LM, Kumar M, Namdeo PK. Sodium Alginate Microspheres for Extending Drug Release: Formulation and *in vitro* Evaluation, International Journal of Drug Delivery,2010;2(1):64-68.
30. Bhusnure Dr Omprakash, Shinde Chandrakant, Gholve Sachin, Shinde Nitin, Sugave R, Rajurkar R, *et al.* Design, Evaluation and Aseptic Refiner Techniques for Microsphere Formation. World Journal of Pharmaceutical Research,2015;4:1870-1902.